CLEAN VERSION OF REPLACEMENT PARAGRAPHS IN THE SPECIFICATION PURSUANT TO 37 C.F.R. §1.121(b)

IN THE SPECIFICATION

On page 96, please delete the paragraph beginning on line 18 and ending on page 97, line 9, and replace with the following paragraph:

These cell lines are used to determine conditions in which a control antisense PKCa phosphorothioate oligonucleotide (GTTCTCGCTGGTGAGTTTCA (SEQ ID NO:1); ISIS3521), included in STEP complexes, results in a decrease in expression of the PKCa-EGFP fusion protein. The efficacy of the oligonucleotide is first confirmed using standard antisense delivery methods (Dean, et al., J Biol Chem 269:16416-24 (1994)) to treat 60 mm dishes of normal HEK-293T cells followed by western blot analysis of PKCa protein levels. PKCa antibodies are commercially available for this purpose (Upstate Biotechnology, Inc.). Following confirmation of the efficacy of the PKCa antisense oligonucleotide, the same twodimensional array analysis of the factors that alter transfection efficiency is employed as was utilized for plasmid DNA transfection (see Preliminary Results and Specific Aim 1A). Basically, the type of cationic lipid and protein included in the DNA complex is varied, as is the ratio of the various DNA complex components. Increased pressure enhances the effect of antisense oligonucleotides following STEP, similar to previous reports that pressure treatment increases the uptake of oligodeoxynucleotides (Mann, et al., Proc Natl Acad Sci U S A 96:6411-6 (1999)). For applying increased pressure, a small Plexiglas chamber with a sealed piston and a pressure gauge is constructed. The chamber is prewarmed to 37°C and filled with 5% CO₂. Each 10 cm tissue culture plate is treated at 1 to 3 atm pressure for 1 to 10 min, and the effect on STEP transfection efficiency is determined as described above.

On page 105, please delete Table 2, and replace with the following Table 2:

Table 2.

Selected Reporter Sequences for Functional Screening of Constitutively Active
Protein Kinases

Reporter/Sequence	Transcription Factor	Reference
AP-I* (TGACTCA) (SEQ ID NO:2)	c-fos, junB, junD	Fisch et al., 1989
CRE* (TGACGTCA) (SEQ ID NO:3)	CREB, CREM, etc.	Benbrook and Jones,
NF-kB*(GGGAATTCC) (SEQ ID NO:4)	NF-kB	1994
SRE* (60 nucleotides)	Elk-l	Lembecher et al.,
p53* (GAAACTGAAACT) (SEQ ID NO:5)	p53	1993
ISRE*(AAACTGAAACTG) (SEQ ID NO:6)	Stat1, Stat2, IRF	Treisman, 1990
GAS*(AGTTTCATATTTACTCTAAATC) (SEQ ID NO:7)	Stat1	Oh et al., 2000
NFAT* (GGAGGAAAAACTGTTCATACAGAAGGCGT) (SEQ ID NO:8)	NF-ATc; NF-ATp	Hiscott et al., 1999
E-box* (CACGTCCACGTC) (SEQ ID NO:9)		Hiscott et al., 1999
E2F* (CTTGGCGGGAGATAGAA) (SEQ ID NO:10)	c-myc	Northrop et al., 1993
pRb* (60 nucleotides)	E2F-1,E2F-2,E2F-3	
Ets-1 (CCAGGAAG) (SEQ ID NO:11)	pRb	Blackwell et al., 199
Oct-1 (ATGCAAATGATAT) (SEQ ID NO:12)	Ets-1	Lam et al., 1995
HNF3(CTAAGTCAATAAT) (SEQ ID NO:13)	Oct-1, Oct-2	Robbins et al., 1990
C/EBPb (tgcagATTGCGCAATctgca) (SEQ ID NO:14)	HNF3	Uchijima et al., 1994
CTF (gccAGCCAATgagcgc) (SEQ ID NO:15)	C/EBPb	Kamps et al., 1990
Egr-1 (CGCCCTCGCCCCGCGCCGGG) (SEQ ID NO:16)	CTF-NF1	Pani et al., 1992
Delta Factor	Egr-1, WT1	Vinson et al., 1993
(CCCCGCTGCCATC) (SEQ ID NO:17)	•	Altman et al., 1994
NF-1	YY-1, F-ACT1, etc.	Cao et al., 1990
(GTTATGGCGACTATCCAGCTTTGTG) (SEQ ID NO:18)		
HSF1 (GAAacCCCtgGAAtaTTcccGAC) (SEQ ID NO:19)	NF-1	Hariharan et al., 199
SIE (TTCCCGTAA) (SEQ ID NO:20)	HSF1	
	Stat1,2,3	Hale and Braithwaite
		1995
		Abravaya et al., 1991
		Boccaccio et al., 199